

Terahertz Time Domain Spectroscopy of Crystalline α -Lactose Monohydrate

Euna Jung¹, Jeonghoi Kim¹, Younho Han¹,
Kiwon Moon¹, Meehyun Lim¹ & Haewook Han¹

¹National Research Laboratory for Nano-THz Photonics,
Department of Electrical and Computer Engineering, Pohang
University of Science and Technology, Pohang 790-784, Korea
Correspondence and requests for materials should be addressed
to H. Han (hhan@postech.ac.kr)

Accepted 24 October 2008

Abstract

Fundamental biological processes require both small- and large-scale structural changes within molecules, including low frequency collective vibrational motions. Many theoretical calculations have predicted that the collective vibrational motion can be characterized as the large-scale intermolecular motions in the terahertz (THz) frequency range. A new measurement technique based on THz time domain spectroscopy (TDS) can probe directly the collective vibrational motions of biomolecules which involve structural changes and specific biological functions. We report the THz TDS of solid state α -lactose monohydrate to characterize the collective vibrational motions. Our measurements show that the THz transmission spectrum of α -lactose monohydrate has strong absorption peaks centered at 0.531, 1.195, 1.38, and 1.81 THz, which result from intermolecular interactions.

Keywords: Terahertz time domain spectroscopy, Low frequency vibrational modes, α -Lactose monohydrate, Intermolecular interactions, Biosensing

Introduction

Terahertz (THz) radiation is an electromagnetic wave in the frequency range of 0.1 to 10 THz between microwave and infrared bands^{1,2}. Due to its non-ionizing and non-invasive characteristics, there has been much attention for various applications of THz radiation, including semiconductor characterization³, medical diagnostics⁴, label-free genetic analysis⁵, and chemical and biological sensing⁶.

The low frequency vibrational and torsional motions of biomolecular systems in condensed phase

depend strongly on intermolecular interactions dominated by hydrogen bonding networks⁷. The theoretical predictions based on simulations of molecular dynamics suggest that the low frequency collective motions of biomolecules occur in THz frequency range⁸⁻¹⁰. Such motions involve the large-scale “breathing” of entire subunits of the macromolecules, which affect the structural changes and functions of biomolecules. The low frequency collective modes are critical for conformational changes in a variety of biological processes, ranging from primary photoisomerization of rhodopsin to enzyme reactions¹¹. The identification of these collective motions can offer the structural and functional information of molecular systems in living cells¹².

THz time domain spectroscopy (TDS) is a new technique to investigate the low frequency molecular dynamics of biomolecular structures in both solid and liquid phases. Many studies have already shown that THz TDS can provide spectroscopic information for a variety of biomolecules such as DNA/RNA^{13,14}, polysaccharides¹⁵, amino acids¹⁶, and proteins¹⁷. In recent years, several research groups have investigated saccharide related molecules at THz frequencies because they are simple enough to study in detail and also of great biological significance^{15,18}. Specifically, α -lactose monohydrate, as shown in Figure 1, is one of the most common disaccharides. In this work, we present our preliminary results on THz TDS of solid state α -lactose monohydrate at 0.2-2.0 THz.

Results and Discussion

The THz pulse signal transmitted through a lactose pellet is shown in Figure 2. The incident THz pulses are delayed due to the refractive index of the samples

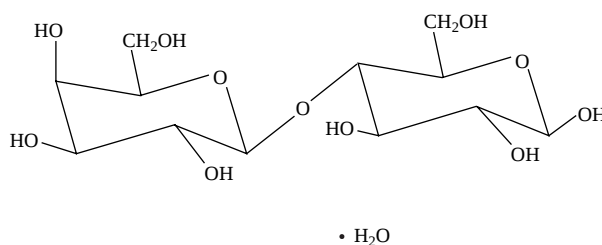


Figure 1. Molecular structure of α -lactose monohydrate.

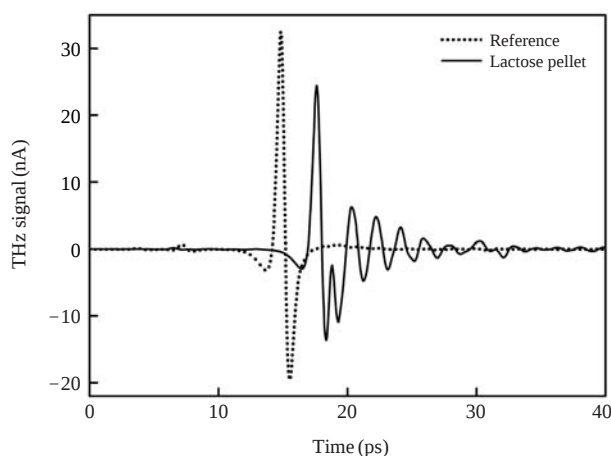


Figure 2. THz signal transmitted through the 1-mm thick lactose pellet. The reference signal was measured without the pellet.

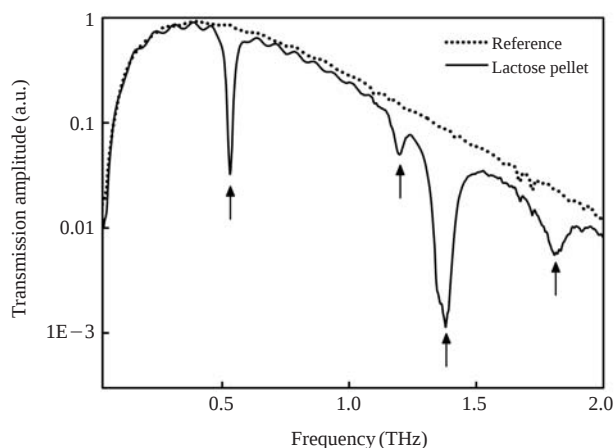


Figure 3. THz Transmission spectrum of α -lactose monohydrate at room temperature.

and attenuated due to absorption and Fresnel loss. The additional small peaks results from the multiple reflections at the two sample-air interfaces and also the dispersion effect due to the complex permittivity of α -lactose monohydrate.

The transmission spectrum of solid state α -lactose monohydrate at room temperature is shown in Figure 3 where the absorption peaks are indicated by arrows. The THz spectrum indicates that α -lactose monohydrate has absorption peaks centered at 0.531, 1.195, 1.38, and 1.81 THz. As reported in a previous work, this strong absorption results from intermolecular vibrations¹⁹.

α -lactose consists of two monosaccharides linked by a glycosidic bond. Density functional theory

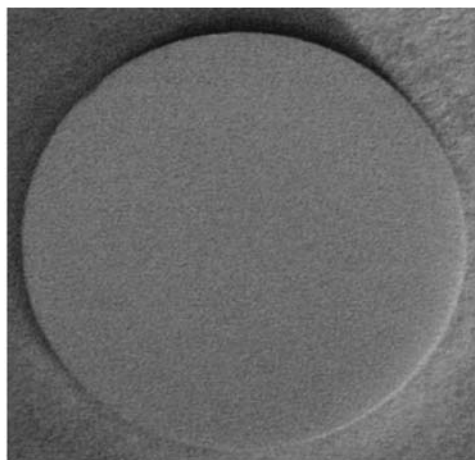


Figure 4. Photograph of α -lactose monohydrate pellet.

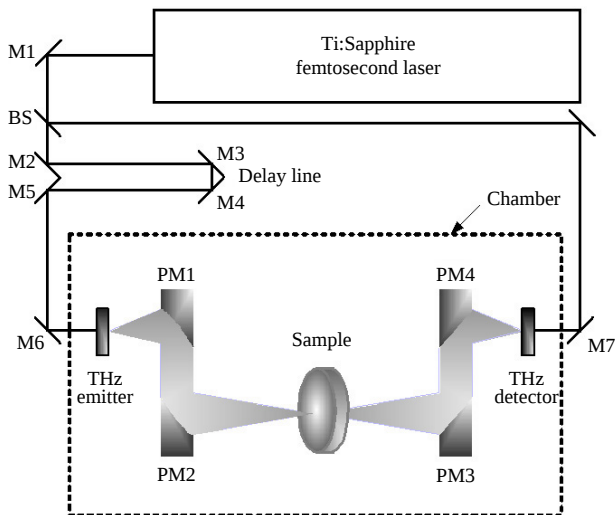


Figure 5. Schematic of THz TDS setup with transmission geometry. M: mirror, BS: beam splitter, PM: off-axis parabolic mirror.

(B3LYP functional) of isolated α -lactose monohydrate predicts intramolecular twisting motion at 0.858 THz²⁰. However, there was no absorption peak around 0.858 THz in our measured spectra, indicating that THz absorption signatures cannot be explained by the intramolecular interactions in an isolated molecule. In addition, a recent study suggests that the lowest frequency THz absorption at 0.531 THz is due to the externally hindered rotational mode with no internal mode contribution²⁰. Our measurements confirm that the low frequency collective motions are mainly affected by intermolecular rather than intramolecular interactions in nature. These collective motions are

associated with interactions between molecules in the hydrogen bonded networks of molecular crystals.

Conclusion

THz absorption spectra of crystalline α -lactose monohydrate with a glycosidic linkage between 0.1 and 2.0 THz were measured by THz TDS. The THz transmission spectra have strong absorption peaks centered at 0.531, 1.195, 1.38, and 1.81 THz. We interpret these THz absorption peaks as the signatures of intermolecular interaction motions of the hydrogen bonded crystalline structure of α -lactose monohydrate. We expect that in near future the THz TDS technique can be applied for biosensors and biochips to identify the spectroscopic fingerprints of specific biomolecules.

Materials and Methods

Sample Preparation

Crystalline spectrophotometric-grade α -Lactose monohydrate powder (Sigma-Aldrich, USA) was used without further purification. To identify the intrinsic spectral features of the biomolecules, the crystalline powder was pressed into 1-mm thick pellets, following standard practice. Figure 4 shows the lactose pellet with 13-mm diameter.

Experimental System

We used a conventional THz TDS system with transmission geometry²¹, shown in Figure 5. The THz pulse was generated by a InAs wafer pumped by a Ti:sapphire laser (Tsunami, Spectra Physics, USA) with a center wavelength of 790 nm, a pulse width of 100 fs, and a repetition rate of 80 MHz. The generated THz pulse was collimated and focused by off-axis parabolic mirrors. The sample was placed at the THz beam waist between two off-axis parabolic mirrors. The transmitted THz signal was detected by a photoconductive antenna fabricated on a low-temperature grown GaAs using the standard optical gating and phase-sensitive detection techniques²². To avoid the absorption due to atmospheric water vapor, the entire THz TDS system was enclosed in a chamber and continuously purged with dry air during the measurements.

Acknowledgements

This study was supported by the Korea Science and

Engineering Foundation through the National Research Laboratory Program (R0A-2005-001-10152-0) funded by the Ministry of Science and Technology, and by the Korean Research Foundation through the Priority Research Centers Program (KRF-2005-005-J13103) and Brain Korea 21 Project funded by the Ministry of Education and Human Resources Development.

References

1. Ferguson, B. & Zhang, X.-C. Materials for terahertz science and technology. *Nature Materials* **1**, 26-33 (2002).
2. Tonouchi, C. Cutting-edge terahertz technology. *Nature Photonics* **1**, 97-105 (2007).
3. Grischkowsky, D., Keiding, S., VanExter, M. & Fattinger, Ch. Far-infrared time-domain spectroscopy with terahertz beams of dielectrics and semiconductors. *J. Opt. Soc. Amer. B* **7**, 2006-2015 (1990).
4. Fitzgerald, A.J. *et al.* Terahertz pulsed imaging of human breast tumors. *Radiology* **239**, 553-540 (2006).
5. Brucherseifer, M. *et al.* Label-free probing of the binding state of DNA by time-domain terahertz sensing. *Appl. Phys. Lett.* **77**, 4049-4051 (2000).
6. Mickan, S.P. *et al.* Label-free bioaffinity detection using terahertz technology. *Phys. Med. Biol.* **47**, 3789-3795 (2002).
7. Fischer, B.M., Walther, M. & Jepsen, P.U. Far-infrared vibrational modes of DNA components studied by terahertz time-domain spectroscopy. *Phys. Med. Biol.* **47**, 3807-3814 (2002).
8. Putnam, B.F., VanZandt, L.L., Prohofsky, E.W. & Mei, W.N. Resonant and localized breathing modes in terminal regions of the DNA double helix. *Bio-phys. J.* **35**, 271-287 (1981).
9. Young, L., Prabhu, V.V. & Prohofsky, E.W. Calculation of far-infrared absorption in polymer DNA. *Phys. Rev. A* **39**, 3173-3180 (1989).
10. Brooks, B. & Karplus, M. Normal modes for specific motions of macromolecules: Application to the hinge bending mode of lysozyme. *Proc. Nat. Acad. Sci. USA.* **82**, 4955-4999 (1985).
11. Balu, R. *et al.* Terahertz spectroscopy of bacteriorhodopsin and rhodopsin: similarities and differences. *Biophys. J.* **94**, 3217-3226 (2008).
12. Berendsen, H.J.C. & Hayward, S. Collective protein dynamics in relation to function. *Curr. Opin. Struct. Biol.* **10**, 165-169 (2000).
13. Markelz, A.G., Roitberg, A. & Heilweil, E.J. Pulsed terahertz spectroscopy of DNA, bovine serum albumin and collagen between 0.1 and 2.0 THz. *Chem. Phys. Lett.* **320**, 42-48 (2000).
14. Fischer, B.M. *et al.* Terahertz time-domain spectroscopy and imaging of artificial RNA. *Opt. Express* **13**, 5205-5215 (2005).
15. Upadhyaya, P.C., Shen, Y.C., Davies, A.G. & Linfield,

- E.H. Far-infrared vibrational modes of polycrystalline saccharides. *Vib. Spectros.* **35**, 139-143 (2004).
16. Ueno, Y., Rungsawang, R., Tomita, I. & Ajito, K. Quantitative measurements of amino acids by terahertz time-domain transmission spectroscopy. *Anal. Chem.* **78**, 5424-5428 (2006).
 17. Xu, J., Plaxco, K.W. & Allen, S.J. Collective dynamics of lysozyme in water: Terahertz absorption spectroscopy and comparison with theory. *J. Phys. Chem. B.* **110**, 24255-24259 (2006).
 18. Heugen, U. *et al.* Solute-induced retardation of water dynamics probed directly by terahertz spectroscopy. *Proc. Nat. Acad. Sci. USA.* **103**, 12301-12306 (2006).
 19. Federici, J.F. *et al.* THz imaging and sensing for security applications-explosives, weapons and drugs. *Semicond. Sci. Technol.* **20**, S266-S280 (2005).
 20. Allis, D.G. *et al.* Assignment of the lowest-lying THz absorption signatures in biotin and lactose monohydrate by solid-state density functional theory. *Chem. Phys. Lett.* **440**, 203-209 (2007).
 21. VanExter, M., Fattinger, Ch. & Grischkowsky, D. Terahertz time domain spectroscopy of water vapor. *Opt. Lett.* **14**, 1128-1130 (1989).
 22. Tani, M., Herrmann, M. & Sakai, K. Generation and detection of terahertz pulsed radiation with photoconductive antennas and its application to imaging. *Meas. Sci. Technol.* **13**, 1739-1745 (2002).